

UNIVERSITY OF MARYLAND CENTER FOR ENVIRONMENTAL SCIENCE  
CHESAPEAKE BIOLOGICAL LABORATORY  
NUTRIENT ANALYTICAL SERVICES LABORATORY  
146 Williams St., Solomons MD 20688  
[www.umces.edu/nutrient-analytical-services-laboratory](http://www.umces.edu/nutrient-analytical-services-laboratory)

**Standard Operating Procedure for  
Determination of Particulate Biogenic Silica in Fresh, Estuarine, and Coastal Waters  
(Reference Method: SM 4500-SiO<sub>2</sub> E-2011)**

**Document #: NASLDoc-036**

**Revision 2025-1  
Replaces revision 2024-1  
Effective May 1, 2025**

**I attest that I have reviewed this standard operating procedure and agree to comply with  
all procedures outlined within this document.**

_____ Employee (Print)	_____ Employee (Signature)	_____ Date
_____ Employee (Print)	_____ Employee (Signature)	_____ Date
_____ Employee (Print)	_____ Employee (Signature)	_____ Date
_____ Employee (Print)	_____ Employee (Signature)	_____ Date

---

Revised by: _____	Date: _____
Reviewed by: _____	Date: _____
Laboratory Supervisor: _____	Date: _____

---

## Changes affecting revision 2025-1

- Section 1.2: Changed MDL from 0.03 to 0.13 mg Si/L
- Section 1.3: Changed Quantitation Limit from 0.09 to 0.39 mg Si/L
- Section 6.1: Added Gallery+ Aqua Master
- Section 7.11: Added wash solution
- Table 2: Removed Dup OR Spike
- Section 11.6: Changed acid from 2.4 mL to 2.0 mL
- Section 12.2: Added Gallery+ Operating Procedure
- Section 12.2.16: Changed ICV from 20.6 to 16.65 mg Si/L
- Section 12.2.17: Changed 10% to 5%

## 1. SCOPE and APPLICATION

- 1.1 The reaction is based on the reduction of silicomolybdate in an acidic solution to heteropoly blue by ascorbic acid. Oxalic acid is added to minimize interference from phosphates. Detection of the silicomolybdate complex is by colorimetry.
- 1.2 A Method Detection Limit (MDL) of 0.13 mg Si/L was determined using the Student's *t* value times the standard deviation of at least seven replicates. Refer to the Student's *t* test table for the appropriate n-1 value.
- 1.3 The Quantitation Limit for biogenic silica was set at 0.39 mg Si/L.
- 1.4 This procedure should be used by analysts experienced in the theory and application of particulate inorganic analysis. Three months of experience with an analyst, experienced in the analysis of biogenic silica, is required.
- 1.5 This method can be used for all programs that require analysis of biogenic silica.
- 1.6 This procedure references SM 4500-SiO<sub>2</sub> E-2011.

## 2. SUMMARY

Particulate samples, collected on a filter pad, are dissolved in NaOH in a 100°C water bath, then cooled in an ice bath to terminate the reaction, and then neutralized with H<sub>2</sub>SO<sub>4</sub> before being analyzed colorimetrically.

## 3. DEFINITIONS

- 3.1 Acceptance Criteria – Specified limits placed on characteristics of an item, process, or service defined in a requirement document. (ASQC)
- 3.2 Accuracy – The degree of agreement between an observed value and an accepted reference value. Accuracy includes a combination of random error (precision) and systematic error (bias) components which are due to sampling and analytical operations; a data quality indicator. (QAMS)
- 3.3 Aliquot – A discrete, measured, representative portion of a sample taken for analysis. (EPA QAD Glossary)
- 3.4 Analytical Range – 0 to 40.3 mg/L Si.
- 3.5 Batch – Environmental samples, which are prepared and/or analyzed together with the same process and personnel, using the same lot(s) of reagents. An **analytical batch** is composed of prepared environmental samples (extracts, digestates, or concentrates) and/or those samples not requiring preparation, which are analyzed together as a group using the same calibration curve or factor. An analytical batch can include samples originating from various environmental matrices and can exceed 20 samples. (NELAC/EPA)
- 3.6 Blank- A sample that has not been exposed to the analyzed sample stream in order to monitor contamination during sampling, transport, storage, or analysis. The blank is subjected to the usual analytical and measurement process to establish a zero baseline or background value and is sometimes used to adjust or correct routine analytical results. (ASQC)

- 3.7 Calibrate- To determine, by measurement or comparison with a standard, the correct value of each scale reading on a meter or other device, or the correct value for each setting of a control knob. The levels of the applied calibration standard should bracket the range of planned or expected sample measurements. (NELAC)
- 3.8 Calibration – The set of operations that establish, under specified conditions, the relationship between values indicated by a measuring device. The levels of the applied calibration standard should bracket the range of planned or expected sample measurements. (NELAC)
- 3.9 Calibration Curve – The graphical relationship between known values, such as concentrations, or a series of calibration standards and their analytical response. (NELAC)
- 3.10 Calibration Method – A defined technical procedure for performing a calibration. (NELAC)
- 3.11 Calibration Standard – A substance or reference material used to calibrate an instrument. (QAMS)
  - 3.11.1 Initial Calibration Standard (STD) – A series of standard solutions used to initially establish instrument calibration responses and develop calibration curves for individual target analytes.
  - 3.11.2 Initial Calibration Verification (ICV) – An individual standard, analyzed initially, prior to any sample analysis, which verifies acceptability of the calibration curve or previously established calibration curve.
  - 3.11.3 Continuing Calibration Verification (CCV) – An individual standard that is analyzed after every 18-23 field sample analysis.
- 3.12 Certified Reference Material (CRM) –A reference material one or more of whose property values are certified by a technically valid procedure, accompanied by or traceable to a certificate or other documentation which is issued by a certifying body. (ISO 17025)
- 3.13 Corrective Action – Action is taken to eliminate the causes of an existing nonconformity, defect, or other undesirable situation in order to prevent a recurrence. (ISO 8402)
- 3.14 Deficiency – An unauthorized deviation from acceptable procedures or practices. (ASQC)
- 3.15 Demonstration of Capability – A procedure to establish the ability of the analyst to generate acceptable accuracy. (NELAC)
- 3.16 Detection Limit – The lowest concentration or amount of the target analyte that can be determined to be different from zero by a single measurement at a stated degree of confidence.
- 3.17 Duplicate Analysis – The analyses of measurements of the variable of interest performed identically on two sub samples (aliquots) of the same sample. The results from duplicate analyses are used to evaluate analytical or measurement precision but not the precision of sampling, preservation, or storage internal to the laboratory. (EPA-QAD)
- 3.18 External Standard (ES) – A pure analyte that is measured in an experiment separate from the experiment used to measure the analyte(s) in the sample.

The signal observed for a known quantity of the pure external standard is used to calibrate the instrument response for the corresponding analyte(s). The instrument response is used to calculate the concentrations of the analyte(s) in the unknown sample.

- 3.19 Field Duplicates (FD1 and FD2) – Two separate samples collected at the same time and place under identical circumstances and treated exactly the same throughout field and laboratory procedures. Analyses of FD1 and FD2 provide a measure of the precision associated with sample collection, preservation, and storage, as well as with laboratory procedures.
- 3.20 Field Reagent Blank (FRB) – A aliquot of reagent water or other blank matrix that is placed in a sample container in the laboratory and treated as a sample in all respects, including shipment to the sampling site, exposure to the sampling site conditions, storage, preservation, and all analytical procedures. The purpose of the FRB is to determine if method analytes or other interferences are present in the field environment.
- 3.21 Holding time – The maximum time that samples may be held prior to analysis and still be considered valid. (40 CFR Part 136) The time elapsed from the time of sampling to the time of extraction or analysis, as appropriate.
- 3.22 Ice Bath – An ice-water mixture used to stop the digestion reaction.
- 3.23 Laboratory Duplicates (LD1 and LD2) – Two aliquots of the same sample taken in the laboratory and analyzed separately with identical procedures. Analyses of LD1 and LD2 indicate precision associated with laboratory procedures, but not with sample collection, preservation, or storage procedures.
- 3.24 Laboratory Reagent Blank (LRB) – A blank matrix (i.e., 10% NaOH) that is treated exactly like a sample including exposure to all glassware, equipment, solvents, and reagents that are used with other samples. The LRB is used to determine if method analytes or other interferences are present in the laboratory environment, the reagents, or the instrument.
- 3.25 Laboratory Control Sample (LCS) – A sample matrix, free from the analytes of interest, spiked with verified known amounts of analytes from a source independent of the calibration standard or a material containing known and verified amounts of analytes. The LCS is generally used to establish intra-laboratory or analyst-specific precision and bias or to assess the performance of all or a portion of the measurement system. (NELAC)
- 3.26 Limit of Detection (LOD) –The lowest concentration level that can be determined by a single analysis and with a defined level of confidence to be statistically different from a blank. Also referred to as MDL. (ACS)
- 3.27 Limit of Quantitation (LOQ) –The minimum levels, concentrations, or quantities of a target variable (target analyte) that can be reported with a specified degree of confidence. The LOQ is set at 3 to 10 times the LOD, depending on the degree of confidence desired. Also referred to as QL.
- 3.28 Linear Dynamic Range (LDR) – The absolute quantity over which the instrument response to an analyte is linear. This specification is also referred to as the Linear Calibration Range (LCR).

- 3.29 Material Safety Data Sheets (MSDS) – Written information provided by vendors concerning a chemical’s toxicity, health hazards, physical properties, fire, and reactivity data including storage, spill, and handling precautions.
- 3.30 May – Denotes permitted action, but not required action. (NELAC)
- 3.31 Method Detection Limit (MDL) – The minimum concentration of an analyte that can be identified, measured, and reported with 99% confidence that the analyte concentration is greater than zero (Standard Methods).
- 3.32 Must – Denotes a requirement that must be met. (Random House College Dictionary)
- 3.33 Photometer – measures the absorbance of the solution in the cell in a multicell cuvette. Light passes from the lamp through the condensing lenses to the interference filter. The plane surface of the first condensing lens is coated with a material that reflects heat and infrared light. The filters are mounted on a filter wheel. There are 15 positions for filters. Each filter corresponds to a wavelength of interest. The 660 nm filter is specified by the test definition for silicate. After passing through the filter the light is converted into a stream of light pulses by a chopper. Then the light is directed via a quartz fiber through a focusing lens and a slit to the beam divider. The beam divider divides the light into two parts. A specified portion is reflected in the reference detector, which monitors the light level fluctuations. The remaining major portion of the light beam goes through the liquid in the cell to the signal detector, which measures the amount of light absorbed.
- 3.34 Precision – The degree to which a set of observations or measurements of the same property, obtained under similar conditions, conform to themselves; a data quality indicator. Precision is usually expressed as standard deviation, variance, or range, in either absolute or relative terms. (NELAC)
- 3.35 Preservation – Refrigeration, freezing, and/or reagents added at the time of sample collection (or later) to maintain the chemical and or biological integrity of the sample.
- 3.36 Quality Control Sample (QCS) –A sample of an analyte of known and certified concentration. The QCS is obtained from a source external to the laboratory and is different from the source of calibration standards. It is used to check laboratory performance with externally prepared test materials. Also referred to as CRM.
- 3.37 Re-Pipette- A calibrated dispenser of reagent to samples.
- 3.38 Run Cycle – Typically a day of operation – the entire analytical sequence from sampling the first standard to the last sample of the day.
- 3.39 Sample Segment – Bar-coded metal tray that holds up to fourteen four milliliter auto analyzer vials containing samples or standards. The user identifies each vial in the operating software.
- 3.40 Sample Segment Holder – An automated temperature-controlled carousel that contains up to six sample segments. This carousel spins in a clockwise or counterclockwise manner to move the sample segments into position for analysis. This carousel format allows for continuous processing.

- 3.41 Sensitivity – The capability of a test method or instrument to discriminate between measurement responses representing different levels (concentrations) of a variable of interest.
- 3.42 Shall – Denotes a requirement that is mandatory whenever the criterion for conformance with the specification requires that there be no deviation. (ANSI)
- 3.43 Should – Denotes a guideline or recommendation whenever noncompliance with the specification is permissible. (ANSI)
- 3.44 Standard Reference Material (SRM) –Material that has been certified for specific analytes by a variety of analytical techniques and/or by numerous laboratories using similar analytical techniques. These may consist of pure chemicals, buffers, or compositional standards. The materials are used as an indication of the accuracy of a specific analytical technique. Also referred to as CRM.
- 3.45 Test Definition – A photometric test consisting of a user-defined testing sequence, reagent additions, calibration standards, incubations, and absorption results.
- 3.46 Test Flow – Functions to define the parameter for reagent and sample dispensing, dilution, incubation, and measurement
- 3.47 Water Bath – A water bath, set to 100 °C, filled with water to a depth that will cover liquid in tubes for digestion.

## **4 INTERFERENCES**











- 4.1 Because both apparatus and reagents may contribute silica, avoid using glassware as much as possible and use reagents low in silica. Phosphate interference can be eliminated by the addition of oxalic acid.
- 4.2 Suspended matter in the sample will scatter light as it passes through the cuvette to the detector. High blank responses will result. Samples may be centrifuged to clarify. The identified sample will be reanalyzed.
- 4.3 Blemishes in the cuvette, as a result of the manufacturing process, will result in high blank responses. The identified sample will be reanalyzed.

## **5 SAFETY**

- 5.1 Safety precautions must be taken when handling reagents, samples, and equipment in the laboratory. Protective clothing including lab coats, safety glasses, and enclosed shoes must always be worn. In certain situations, it will be necessary to also use gloves and/or a face shield. If solutions come in contact with eyes, flush with water continuously for 15 minutes. If solutions come in contact with skin, wash thoroughly with soap and water. Contact Solomons Rescue Squad (911) if emergency treatment is needed and also inform the Chesapeake Biological Laboratory (CBL) Associate Director of Administration and Facilities Maintenance of the incident. Contact the Chesapeake Biological Laboratory (CBL) Associate Director of Administration and Facilities Maintenance if additional treatment is required.

- 5.2 The toxicity or carcinogenicity of each reagent used in this procedure may not have been fully established. Each chemical should be regarded as a potential health hazard and exposure should be as low as reasonably achievable. Cautions are included for known hazardous materials and procedures.
- 5.3 Do not wear jewelry when troubleshooting electrical components. Even low voltage points are dangerous and can injure if allowed to short circuit.
- 5.4 The following hazard classifications are listed for the chemicals used in this procedure. Detailed information is provided on Material Safety Data Sheets (MSDS).

Table 1:

Chemical	Health Hazard	Fire Hazard	Instability Hazard	Specific Hazard	
Sodium Hydroxide	3	0	1	ALK, COR	 
Sulfuric acid	4	0	2	ACID, COR	  
Oxalic acid	3	0	0	ACID, COR	 
Ascorbic acid	1	0	0	ACID	
Ammonium molybdate	2	0	0	IRRITANT	 
Sodium silicofluoride	2	0	0	IRRITANT	
Sodium Hypochlorite	3	0	0		
On a scale of 0 to 4 the substance is rated on four hazard categories: health, flammability, reactivity, and contact. (0 is non-hazardous and 4 is extremely hazardous)					
<p><b>HAZARD RATING</b></p> <p>Health Hazard - Blue: 4 – deadly, 3 – extreme danger, 2 – hazardous, 1 – slightly hazardous, 0 – normal material</p> <p>Fire Hazard - Red: Flash Points: 4 – below 73° F, 3 – below 100° F, 2 – below 200° F, 1 – above 200° F, 0 – will not burn</p>					



Instability Hazard - Yellow: 4 – may detonate, 3 – Shock and heat may detonate, 2 – violent chemical change, 1 – unstable is heated, 0 - stable

Specific Hazard - White: Acid = ACID, Alkali = ALK, Corrosive = COR, Oxidizer = OXY

## 6 EQUIPMENT AND SUPPLIES

- 6.1 Aquakem 250 or Gallery+ Aqua Master multi-wavelength automated discrete photometric analyzer. Control software operates on a computer running Microsoft Windows 7 or a newer operating system.
- 6.2 Refrigerator, capable of maintaining  $4 \pm 2^{\circ}\text{C}$ .
- 6.3 Freezer, capable of maintaining  $-20 \pm 5^{\circ}\text{C}$ .
- 6.4  $100^{\circ}\text{C}$  hot water bath.
- 6.5 Ice water bath
- 6.6 Lab ware – All reusable labware (NO glass, use polypropylene) should be sufficiently clean for the task objectives. This laboratory cleans all labware related to this method with a 10% NaOH (w/v) rinse.
- 6.7 2 digital timers
- 6.8 Polypropylene centrifuge tubes with caps, and racks
- 6.9 2 polypropylene re-pipettes
- 6.10 A centrifuge

## 7 REAGENTS AND STANDARDS

- 7.1 Purity of Water – Unless otherwise indicated, references to water shall be understood to mean reagent water conforming to Specification D 1193, Type I. Freshly prepared water should be used for making the standards intended for calibration. The detection limits of this method will be limited by the purity of the water and reagents used to make the standards.
- 7.2 Purity of Reagents – Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available. Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without compromising the accuracy of the determination.
- 7.3 Sodium Hydroxide, 0.2 N-

Sodium Hydroxide (NaOH) pellets	8g
Reagent Water	up to 1000mL

In a 1000mL plastic volumetric flask, dissolve 8g of Sodium hydroxide pellets in ~400mL reagent water and dilute to 1000mL with reagent water. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number and balance ID number in the Analytical Reagent log book. Store the flask at room temperature in the dark and make every 3 months.
- 7.4 Sulfuric Acid, 1N-

Sulfuric acid ( $\text{H}_2\text{SO}_4$ ), concentrated (sp. Gr. 1.84) 28mL

Reagent Water up to 1000mL

In a 1000mL plastic volumetric flask, add 28mL of Sulfuric acid conc. to ~400mL reagent water and dilute to 1000mL with reagent water. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number in the Analytical Reagent log book. Store the flask at room temperature in the dark and make every 3 months.

7.5 Oxalic Acid Solution -

Oxalic acid ( $\text{H}_2\text{C}_2\text{O}_4 \cdot 2\text{H}_2\text{O}$ ) 100g

Reagent water up to 1000mL

In a 1000mL plastic volumetric flask, dissolve 100g of oxalic acid in ~400mL reagent water and dilute to 1000mL with reagent water. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number and balance ID number in the Analytical Reagent log book. Store the flask at room temperature in the dark and make every 12 months.

7.6 Ascorbic Acid Solution -

Oxalic acid ( $\text{H}_2\text{C}_2\text{O}_4 \cdot 2\text{H}_2\text{O}$ ) 1.25g

Ascorbic acid ( $\text{C}_6\text{H}_8\text{O}_6$ ), U.S.P. quality 25g

Reagent water up to 250mL

In a 250mL plastic volumetric flask, dissolve 1.25g of oxalic acid in ~100mL of reagent water. Add 25g of ascorbic acid and mix until dissolved. Dilute to 250mL with reagent water. Write name of preparer, preparation date, reagent manufacturers, manufacturers' lot numbers and balance ID number in the Analytical Reagent log book. Store at 4°C and make every 4 months.

7.7 Ammonium Molybdate Solution -

Ammonium molybdate [ $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}$ ] 3.0g

Reagent water up to 100mL

In a 100mL plastic volumetric flask, dissolve 3.0g ammonium molybdate in ~80mL of reagent water. Dilute to 100mL with reagent water. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number and balance ID number in the Analytical Reagent log book. Store in the dark at room temperature. Reagent is stable for 48 hours.

7.9 Sulfuric Acid Solution, 0.7N -

Sulfuric acid ( $\text{H}_2\text{SO}_4$ ), concentrated (sp. Gr. 1.84) 4.0mL

Reagent water up to 1000mL

In a 1000mL plastic volumetric flask, add 4.0 mL of concentrated sulfuric acid and 21.4mL of stock phosphate solution to ~300mL of reagent water. Dilute to 1000mL with reagent water. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number in the Analytical Reagent log book. Store at 4°C and make every 6 months.

7.10 Stock Silicate Standard, 10,000 $\mu\text{M}$

Sodium silicofluoride ( $\text{Na}_2\text{SiF}_6$ ), dried at 45°C 1.88g

Reagent water up to 1000mL

In a 1000mL plastic volumetric flask, dissolve 1.88g of sodium silicofluoride in ~400mL of reagent water. Dilute to 1000mL with reagent water (1ml contains 10 $\mu\text{moles}$  Si). Write name of preparer, preparation date, standard manufacturer,

manufacturer lot number and balance ID number in the Analytical Standard log book.  
Store in a plastic container at room temperature. Make fresh every 6 months.

#### 7.11 Cleaning/ Wash Solution –

Bleach	55.0 mL
--------	---------

In a 100 mL volumetric flask, dilute 55.0 mL of bleach to volume with 45mL reagent water to yield a concentration of 55% (v/v) bleach. Write name of preparer, preparation date, reagent manufacturer, manufacturer lot number in the Analytical Reagent log book. Reagent is stable for 6 months.

#### 7.12 Standards

##### 7.12.1 Stock Silica Standard

Sodium silicofluoride ( $\text{Na}_2\text{SiF}_6$ ), dried at 45 °C, 1.88g  
Deionized water up to 1L

##### 7.12.2 Secondary Silica Standard

Stock Silica Standard	25 mL
Deionized water up to	100 mL

##### 7.12.3 Working Silica Standards

To labeled 50 mL polypropylene screw cap centrifuge tubes, add 0, 0.1, 0.2, 0.3, 0.4, 0.5 and 0.6 mL of Secondary Silica Standard. Digest standards as samples. After digestion, standard tubes will contain 0, 6.94, 13.8, 20.6, 27.2, 33.75 and 40.3 mg Si/L as Si, respectively.

## 8 SAMPLE COLLECTION, PRESERVATION, AND STORAGE

8.1 Filter a known volume (volume dependent on water source) of water through a 0.4  $\mu\text{m}$  Nucleopore™ polycarbonate filter.

8.2 Fold filter in half, sample inside, and place in a labeled 50 ml polypropylene centrifuge tube, cap, and freeze at -20°C.

## 9 QUALITY CONTROL

9.1 The laboratory is required to operate a formal quality control (QC) program. The minimum requirements of this program consist of an initial demonstration of laboratory capability and the continued analysis of laboratory instrument blanks and calibration standard material, analyzed as samples, as a continuing check on performance. The laboratory is required to maintain performance records that define the quality of data generated.

#### 9.2 Initial Demonstration of Capability

9.2.1 The initial demonstration of capability (iDOC) - is used to characterize instrument performance (MDLs) and laboratory performance (analysis of QC samples) prior to the analyses conducted by this procedure.

9.2.2 Quality Control Sample (QCS/SRM) – When using this procedure, a quality control sample is required to be analyzed at the beginning and end of the run, to verify data quality and acceptable instrument performance. If the determined concentrations are not within  $\pm 10\%$  of the certified values, performance of the determinative step of the method is unacceptable. The source of the problem must be identified and corrected

before either proceeding with the initial determination of MDLs or continuing with analyses.

- 9.2.3 Method Detection Limits (MDLs) – MDLs should be established for biogenic silica using a low-level natural water sample, typically three to five times higher than the estimated MDL. To determine the MDL values, analyze at least seven replicate aliquots of water. Perform all calculations defined in the procedure (Section 12) and report the concentration values in the appropriate units. Calculate the MDL as follows:

$$\text{MDL} = St_{(n-1, 1-\alpha=0.99)}$$

Where,

S = Standard deviation of the replicate analyses.

n=number of replicates

$t_{(n-1, 1-\alpha=0.99)}$  = Student's *t* value for the 99% confidence level with n-1 degrees of freedom

MDLs shall be determined yearly and whenever there is a significant change in instrument response or a significant change in instrument configuration.

### 9.3 Assessing Laboratory Performance

- 9.2.1 Laboratory Reagent Blank (LRB) – The laboratory must analyze at least one LRB with each batch of samples. For this analysis, a blank filter is treated the same as a sample and used as the LRB. Analyte found in LRB indicates possible reagent or laboratory environment contamination. LRB data are used to assess and correct contamination from the laboratory environment.
- 9.2.2 Quality Control Sample (QCS)/ Standard Reference Material (SRM) – When using this procedure, a quality control sample is required to be analyzed at the beginning of the run and end of the run, to verify data quality and acceptable instrument performance. If the determined concentrations are not within  $\pm 3s$  of the certified values, the performance of the determinative step of the method is unacceptable. The source of the problem must be identified and corrected before either proceeding with the initial determination of MDLs or continuing with the analyses. The results of these QCS/SRM samples shall be used to determine batch acceptance.
- 9.2.3 The QCS are obtained from a source external to the laboratory and different from the source of calibration standards.
- 9.2.4 Control Charts – The Accuracy Control Chart for QCS/SRM samples and reagent blanks is constructed from the average and standard deviation of the 20 most recent QCS/SRM measurements. The accuracy chart includes upper and lower warning levels (WL =  $\pm 2s$ ) and upper and lower control levels (CL =  $\pm 3s$ ). These values are derived from stated values of the QCS/SRM. The standard deviation (*s*) is specified relative to statistical confidence levels of 95% for WLs and 99% for CLs. Enter QCS/SRM results on the chart each time the sample is analyzed
- 9.2.5 Calibration Verification, Initial and Continuing (ICV/CCV)– Immediately following calibration (ICV) and following every 18-23 samples (CCV), one calibration verification of a duplicate standard is analyzed to assess

instrument performance. The CCVs are made from the same material as calibration standards and are to be within  $\pm 3s$  of the expected value. Failure to meet the criteria requires correcting the problem, including reanalysis of any affected samples. If not enough sample exists, the data must be qualified if reported.

#### 9.4 Assessing Analyte Recovery

9.2.1 Analyte recovery is assessed through percent recoveries of laboratory spikes.

9.2.2 Percent Recovery = (Actual Value/ Expected Value) X 100

#### 9.5 Assessing Analyte Precision – Relative Percent Difference

9.2.1 Analyte replication is assessed through duplicate analyses of samples – Relative Percent Difference.

9.2.2  $RPD = \frac{\text{Laboratory Duplicate Result 1} - \text{Laboratory Duplicate Result 2}}{[(\text{Laboratory Duplicate Result 1} + \text{Laboratory Duplicate Result 2})/2]} \times 100$

#### 9.6 Corrective Actions for Out of Control Data

9.2.1 Control limit – If one measurement exceeds Accuracy Control Chart CL, repeat the analysis immediately. If the repeat measurement is within the CL, continue analyses; if it exceeds the CL, discontinue analyses and correct the problem.

9.2.2 Warning limit – If two out of three successive points exceed Accuracy Control Chart WL, analyze another sample. If the next point is within WL, continue analyses; if the next point exceeds the WL, evaluate potential bias and correct the problem.

9.2.3 Trending – If seven successive Accuracy Control Chart measurements are on the same side of the central line, discontinue analyses and correct the problem.

9.2.4 When external QCS samples are out of control, correct the problem. Reanalyze the samples analyzed between the last in-control measurement and the out-of-control one.

9.2.5 When external CCV samples are out of control, correct the problem. Reanalyze the samples analyzed between the last in-control measurement and the out-of-control one.

9.7 General Operation - To assure optimal operation and analytical results, the Reagent Blank and CCV are tracked daily in the raw data file, copied to Reagent Blank and CCV Control Charts.

Table 2:

QC Indicator	Acceptance/ Action Limits	Action	Frequency (Batch)
Correlation Coefficient	$\geq 0.995$	If $<0.995$ , evaluate data points of the calibration curve. If any data point is outside established limits, reject as outlier.	1 per batch if acceptable.
Quality Control Sample (QCS)/ Certified Reference Material (CRM)	$\pm 10\%$	If QCS value is outside $\pm 10\%$ of the target value reject the run, correct the problem and rerun samples.	Beginning of run and at end of run.
Initial Calibration Verification (ICV)	$\pm 10\%$	Recalibrate if outside acceptance limits.	Beginning of run following standard curve.
Continuing Calibration Verification (CCV)	$\pm 10\%$	If outside $\pm 10\%$ , correct the problem. Rerun all samples following the last in-control CCV.	After every 18-23 samples.
Method Blank/Laboratory Reagent Blank (LRB)	$\leq$ Method Quantitation Limit	If the LRB exceeds the quantitation limit, results are suspect. Rerun the LRB. If the concentration still exceeds the quantitation limit, reject or qualify the data, or raise the quantitation limit.	Following the ICV and after every 18-23 samples following the CCV.
Laboratory Fortified Sample Matrix Spike	$\pm 20\%$	If the recovery of any analyte falls outside the designated acceptance limits and the QCS is in control, the recovery problem is judged matrix induced. Repeat the LFM and if the sample results are again outside the acceptable recovery range, the sample should be reported with a "matrix induced bias" qualifier.	1/20
Laboratory Duplicate	$\pm 20\%$	If the RPD fails to meet the acceptance limits, the samples should be reanalyzed. If the RPD again fails to meet the acceptance limits, the sample must be reported with a qualifier identifying the	1/20

		sample analysis result as not having acceptable RPD for duplicate analysis.	
--	--	---	--

## 10 CALIBRATION AND STANDARDIZATION

- 10.1 Calibration – Daily calibration must be performed before sample analysis may begin. Seven-point calibrations are used. The reagent blank with no Si added is used as the zero standard.
- 10.2 Working Silica Standards: To labeled 50 mL polypropylene screw cap centrifuge tubes, add 0, 0.1, 0.2, 0.3, 0.4, 0.5 and 0.6 mL of Secondary Silica Standard. Digest standards as samples. After digestion, standard tubes will contain 0, 6.94, 13.8, 20.6, 27.2, 33.75 and 40.3 µg Si, respectively.
- 10.3 The instrument software prepares a standard curve for each set of calibrators. A graph plotting measured absorbance against standard concentration is presented for review and approval. If acceptance criteria are not met the entire curve can be reanalyzed or individual standards can be reanalyzed. One standard value (original or reanalyzed) for each and every standard is incorporated in the curve. The coefficient of determination (Pearson's r value) for the calibration curve as well as the calculated concentration of each calibrator is reviewed. The calculated value of each calibrator must be within ten percent of the expected value. The coefficient of determination (Pearson's r value) for the calibration curve must be greater than 0.995.

## 11 PROCEDURE FOR SAMPLE PREPARATION

- 11.1 Fill water bath to a depth that will cover liquid in tubes and heat to 100 °C.
- 11.2 Prepare an ice water bath.
- 11.3 To the sample pad in the centrifuge tube, add 10 ml of 0.2 N Sodium Hydroxide from a polypropylene re-pipette. Make sure that the filter pad is covered by Sodium Hydroxide. Cap the tube leaving it loosened ¼ turn.
- 11.4 Place the centrifuge tube in the 100 °C water bath for exactly 20 minutes.
- 11.5 After exactly 20 minutes, remove the tube from the hot water bath and place in ice water bath for exactly 4 minutes.
- 11.6 After 4 minutes remove tube from ice bath and add 2.0 ml of 1 N Sulfuric Acid Solution to tube to neutralize. Cap and shake. The samples can now be stored at room temperature until analyzed.

## 12 PROCEDURE –

### 12.1 GALLERY+ DAILY OPERATIONS AND QUALITY CONTROL

- 12.1.1. Turn on the computer. Enter Windows user name and password. Open Gallery Discrete Analyzer software and enter software username and password. Once software is running, turn on instrument and allow connection between instrument and computer to complete.

- 12.1.2. Discard any water remaining in the water reservoir from the previous analytical run. Fill the water reservoir with fresh reagent water. Connect the water line to the reservoir and slide the reservoir into the right slot at the bottom of the instrument.
- 12.1.3. Remove from refrigerator samples and SRM that will be analyzed that day. Begin daily bench sheet documentation.
- 12.1.4. Place cuvette waste container into the center slot at the bottom of the instrument. Connect the waste line and insert the waste container into the left slot at the bottom of the instrument.
- 12.1.5. In the instrument software select F5 tab, 2 Actions, and Extra Wash. – complete at least five extra wash cycles.
- 12.1.6. Perform Start Up operations by selecting Start Up at the bottom of the F1 Home Page.
- 12.1.7. Gather working standards and reagents from their appropriate storage area during startup. Assess standards and reagents. Remake anything that has exceeded the time over which it is considered stable.
- 12.1.8. Once startup is complete, check the instrument water blanks by selecting F5 tab, 6 Water Blank Results. If any of the instrument blanks are outside their predefined and software-controlled limits, the user will be notified on the Home page. User takes corrective action to return instrument functions to controlled limits.
- 12.1.9. Load reagents, select F3 tab, 2 Reagents, select an open box, insert new vial, select the appropriate reagent from list. Load reagents into reagent carousel inside the refrigerated reagent compartment located under the right panel.
- 12.1.10. Transfer extract to an AutoAnalyzer cup using a polyethylene Pasteur pipette. Avoid particulate pieces. Samples should be centrifuged at 1700 rpm for 10 minutes.
- 12.1.11. Load working standards and quality control standards into a sample segment. Identify the standards in their positions by selecting F2 tab, 3 racks, select rack, Add sample, select calibrator and controls, select the corresponding calibrator and control ID. Insert the rack into the sample carousel located under the left side panel.
- 12.1.12. Select the methods to be calibrated by selecting F4 tab, 1 Cal/QC selection, select calibration tab. Select BiSi as the method to be calibrated, confirm that the appropriate calibration standards, check standards, and QC standards are loaded for the calibration. After selecting the method select Calibrate at the bottom of the page. The method will now show as pending. Return to the main page.
- 12.1.13. Begin calibration from the F1 home page, select Start, Begin Calibration. See test flow below for stepwise instrument functions for the analysis of standards and samples.

Test Flow – Method of Analysis, Stepwise

- 100 µL SAMPLE to cuvette
- End point absorbance measurement at 660 nm for sample blank determination
- 31 µL sulfuric acid solution (H<sub>2</sub>S Si) to cuvette with mixing



- 39  $\mu$ L ammonium molybdate solution (MOL Si) reagent to cuvette with mixing
- Incubation, 30 seconds
- 62  $\mu$ L oxalic acid solution (OXA Si) to cuvette with mixing
- Incubation, 30 seconds
- 16  $\mu$ L ascorbic acid solution (ASC Si) to cuvette with mixing
- Incubation, 600 seconds
- End point absorbance measurement, 660 nm
- Software processes absorbance value and uses calibration curve to calculate analyte concentration ( $\mu$ g/L of Si)
- User is notified if any measured values used to calculate final concentration are outside preset limits. If so, user has options to accept result, rerun the sample or rerun the sample diluted to a user or software specified factor.

12.1.14. Organize samples and reagent blanks while instrument performs calibrations.

12.1.15. As calibration curves are produced by the instrument, review them for acceptability. The instrument software prepares a standard curve for each set of calibrators. A graph plotting measured absorbance against standard concentration is presented for review and approval. If acceptance criteria are not met, either the entire curve shall be reanalyzed or individual standards shall be reanalyzed, depending on the violation. One standard value (original or reanalyzed) for each and every calibrator is incorporated in the curve. Select F4 tab, 3 Calibration Results, Accept, Rerun, or Reject as needed.

12.1.16. Once calibration curves are accepted, samples are loaded into the sample segments and segments are loaded into the instrument for analysis. Load samples select F2 tab, 3 Racks, select segment, Add sample, Fill Rack, select sample, enter sample ID.

12.1.17. Analysis can be requested two ways. Add analysis requests to racks, select F2 tab, 3 Racks, Request to rack, select test from Bi-Z list, select Save. OR. Add analysis request to individual samples, select F2 tab, 1 samples, select sample from list, select test from Bi-Z list, select Save.

12.1.18. Quality Control samples are pre-programmed in the method to be analyzed, following Table 2. The first samples analyzed should be ICV (initial calibration verification) samples. There should be one sample for each calibration curve, of a concentration close to the middle of each range. The following are ICV samples for the curve: CRM 16.65 mg Si/L for BiSi. CCV (Continuing Calibration Verification) (20.6 mg Si/L standard) follows every 10 samples. Standard Reference Material (SRM) are analyzed after every 10 samples, following CCV. Throughout the analytical batch, samples are chosen as laboratory duplicates and laboratory spikes to assess analyte precision and analyte recovery, respectively. The total number of duplicates and spikes performed will be equal to or greater than five percent of the total number of samples in the analytical batch.

- 12.1.19. As sample analysis is complete, results must be reviewed and accepted manually. If results fall outside acceptance limits, the sample should be reanalyzed. If sample result exceeds the highest standard of the highest calibration range, the samples can be automatically diluted by the instrument and reanalyzed. If the result is such that it will fall within a higher calibration range, it should be reanalyzed in the higher range. If the result is such that it will fall within a lower calibration range, it should be reanalyzed within the lower range. Result can be accessed by selecting F2 tab, 4 Results, Accept, Reject, and Rerun as needed.
- 12.1.20. Upon completion of all analysis, results should be saved to a daily report file. Select F2 tab, 5 Reports, All results to file. The file is then named by the run date. The daily report file for analytical batch of January 2, 2017 would be named 010217. The file is converted to Microsoft Excel for data work up and copied to a removable flash drive. Remaining samples are discarded.
- 12.1.21. All reagents are removed from the reagent chamber and returned to their appropriate storage areas. Reagents that have exceeded their stability period are discarded.
- 12.1.22. Initiate the shutdown procedure. Insert wash solution, select F2 tab, 3 racks, select rack, Add sample, select controls, select WashSol. Clear Daily Files, select F5 tab, 2 Actions, select Clear Daily Files. Initiate Standby from the home tab F1 select standby at the bottom of the screen. Once Standby is complete, remove the Wash Solution from the instrument. Exit the software, select F5 tab, 2 Actions, Exit, select Yes. Turn off instrument and computer.
- 12.1.23. The waste is flushed down the drain with copious amounts of tap water. The waste cuvette box is moved to the fume hood and covered. The instrument is wiped clean of drips or splashes.

## **12.2. AQUAKEM 250 DAILY OPERATIONS AND QUALITY CONTROL**

- 12.2.1 Turn on computer. Computer will automatically initiate Konelab software. Once software is running, turn on instrument and allow connection between instrument and computer to complete.
- 12.2.2 Discard any water remaining in the water reservoir from the previous analytical run. Fill the reservoir with fresh reagent water.
- 12.2.3 Begin daily bench sheet documentation.
- 12.2.4 Place cuvette waste box into cuvette waste sliding drawer.
- 12.2.5 Once water reservoir is full, use instrument software to click More, Instrument Actions, and Perform Water Wash. – complete at least five water wash cycles.
- 12.2.6 After performing water washes, clean the dispensing needle by performing test washes. Click More, Instrument Actions, More, Adjustment Program. Once in the Adjustment Program click, 4-Dispensing Unit, 1-Dispenser, 8-Test Wash. Perform 8 to 10 Test Washing. When complete, press “Q” to quit until you are able to bring up the Main Page.
- 12.2.7 Perform Start Up operations by clicking Start Up at the bottom of the Main Page.

- 12.2.8 Gather reagents from their appropriate storage areas during startup. Assess standards and reagents. Remake anything that has exceeded the time over which it is considered stable.
- 12.2.9 Once startup is complete, check the instrument water blanks by clicking More, Instrument Actions, More, Check Water Blank. If any of the instrument blanks are outside their predefined and software-controlled limits, the user will be notified on the main menu page. User takes corrective action to return instrument functions to controlled limits.
- 12.2.10 Load reagents into reagent carousel and place into refrigerated reagent compartment. Reagent position can be found by clicking reagents at the top of the main page.
- 12.2.11 To analyze, transfer extract to an AutoAnalyzer cup using a polyethylene Pasteur pipette. Avoid particulate pieces. Samples should be centrifuged at 1700 rpm for 10 minutes.
- 12.2.12 Load working standards into a sample segment, identify the standards in their positions from the drop-down menus at the individual segment positions, and load them into the instrument. (Click Samples from the top of the Main Page, then click desired segment number).
- 12.2.13 Select the methods to be calibrated by clicking Calibr. /QC Selection on the bottom of the main page. Click BISICBL as the method to be calibrated, and then click Calibrate at the bottom of the page. The method will now show as pending. Return to the main page.
- 12.2.14 Begin calibration by clicking Page Up on the keyboard. This may also be a green button on the keyboard– See test flow below for stepwise instrument functions for the analysis of standards and samples.

Test Flow – Method of Analysis, Stepwise

- 100 µL SAMPLE to cuvette
- End point absorbance measurement at 660 nm for sample blank determination
- 31 µL sulfuric acid solution (H2S SILCBL) to cuvette with mixing
- 39 µL ammonium molybdate solution (MOL SILCBL) reagent to cuvette with mixing
- Incubation, 30 seconds
- 62 µL oxalic acid solution (OXA SILCBL) to cuvette with mixing
- Incubation, 30 seconds
- 16 µL ascorbic acid solution (ASC SILCBL) to cuvette with mixing
- Incubation, 600 seconds
- End point absorbance measurement, 660 nm
- Software processes absorbance value and uses calibration curve to calculate analyte concentration (µg/L of Si)
- User is notified if any measured values used to calculate final concentration are outside preset limits. If so, user has options to accept result, rerun the sample or rerun the sample diluted to a user or software specified factor.

- 12.2.15 As calibration are produced by the instrument, review them for acceptability. The instrument software prepares a standard curve for each

set of calibrators. A graph plotting measured absorbance against standard concentration is presented for review and approval. If acceptance criteria are not met, either the entire curve shall be reanalyzed or individual standards shall be reanalyzed, depending on the violation. One standard value (original or reanalyzed) for each and every calibrator is incorporated in the curve.

- 12.2.16 Once calibration curves are accepted, samples are loaded into the sample segments and loaded into the instrument for analysis. After the reagent blanks the first samples analyzed should be ICV (initial calibration verification) samples. There should be one sample for each calibration curve, of a concentration close to the middle of the range. The recommended ICV for this curve is 16.65 mg Si/L.
- 12.2.17 Samples are loaded into the segments and analyzed. Run CCV (Continuing Calibration Verification) samples follow every 18-23 samples. Standard Reference Material (SRM) samples as well as Laboratory Reagent Blanks (LRB) are scattered throughout the analytical batch. Throughout the analytical batch, samples are chosen as laboratory duplicates and laboratory spikes to assess analyte precision and analyte recovery, respectively. The total number of duplicates and spikes performed will be equal to or greater than five percent of the total number of samples in the analytical batch.
- 12.2.18 As sample analysis is complete, results must be reviewed and accepted manually. If results fall outside acceptance limits, the sample should be reanalyzed. If sample result exceeds the highest standard of the calibration range, the samples can be automatically diluted by the instrument and reanalyzed.
- 12.2.19 Upon completion of all analysis, results should be saved to a daily report file. Click Report on the bottom of the main page, More, Results To File, and select one row per result. The file is then named by the run date. The daily report file for analytical batch of January 2, 2017 would be named 010217. The file is converted to Microsoft Excel for data work up and copied to a removable flash drive. Remaining samples are discarded.
- 12.2.20 All reagents are removed from the reagent chamber and returned to their appropriate storage areas. Reagents that have exceeded their stability period are discarded.
- 12.2.21 Initiate the shutdown procedure, click on Stand By at the bottom of the main page and insert AquaKem Cleaning Solution into the instrument. Daily files are cleared from the instrument software by clicking More, Management, Clear Daily Files. Once prompted remove the AquaKem Cleaning Solution from the instrument. The software is exited and the instrument is turned off. The computer is turned off.
- 12.2.22 The waste is flushed down the drain with copious amounts of tap water. The waste cuvette box is moved to the fume hood and covered. The instrument is wiped clean of drips or splashes.

## **13 DATA ANALYSIS AND CALCULATIONS**

- 13.1 Upon completion of all analysis, results are saved to a daily report file. The file is named by the run date. The daily report file for the analytical batch of January 1, 2017, would be named 010117. Raw results for each run are copied into a Microsoft Excel spreadsheet. Data are sorted by sample name and time of analysis so that all samples will be displayed by number and results for each sample will be displayed consecutively.
- 13.2 Dilution by the instrument is noted by software as analysis ensues and, is also, documented in the data report spreadsheet. The analyst examines each row of data. Results are eliminated that are outside the limits of the calibration range or have an unrepeated high blank response greater than 0.001 absorbance units.

13.3 Calculation for  $\mu\text{g Si/L}$

$$\text{Corrected } \mu\text{g Si/L} = \frac{(\text{Sample Value } (\mu\text{g Si/L}) - \text{Blank Value } (\mu\text{g Si/L}))}{\text{Volume filtered (L)}}$$

13.4 Calculation for  $\text{mg Si/L}$

$$\text{mg Si/L} = \frac{\text{Corrected } \mu\text{g Si/L}}{1000}$$

13.5 Spike Calculation

$$\text{Actual Spike Value} = \text{Sample Value} - \text{Blank Value}$$

$$\text{Original Sample Value} = \text{Sample Value} - \text{Blank Value}$$

$$\text{Expected Spike Value} = \frac{(\text{Original Sample Value} + 2.06)}{1.1}$$

## 14 WASTE MANAGEMENT AND POLLUTION

- 14.1 Pollution prevention encompasses any technique that reduces or eliminates the quantity of toxicity of waste at the point of generation. Numerous opportunities for pollution prevention exist in laboratory operations. The USEPA has established a preferred hierarchy of environmental management techniques that places pollution as the management option of first choice. Whenever feasible, laboratory personnel should use pollution prevention techniques to address their waste generation. When wastes cannot be feasibly reduced at the source, the agency recommends recycling as the next best option.
- 14.2 For information about pollution prevention that may be applicable to laboratories and research institutions, consult “Less is Better: Laboratory Chemical Management for Waste Reduction”, available from the American Chemical Society, Department of Government Relations and Science Policy, 1155 16<sup>th</sup> Street N. W., Washington, D.C. 20036.

## 15 REFERENCES

- 15.1 U.S. Environmental Protection Agency, 1997. Methods for the Determination of Chemical Substances in Marine and Estuarine Environmental Samples. Method 366.0. U.S. Environmental Protection Agency. Washington, D.C.

- 15.2 Paasche, E. 1973. Silicon and the ecology of marine plankton diatoms. 1. *Thalassiosira pseudonana* (*Cyclotella nana*) grown in a chemostat with silicate as limiting nutrient. Mar. Biol. 19:117-126.
- 15.3 Technicon Industrial Method No. 811-86T. November 1986. Technicon Industrial Systems. Tarrytown, New York, 10591.
- 15.4 APHA, Standard Methods for the Examination of Water and Wastewater, Method #4500-SiO<sub>2</sub> E-2011, 23<sup>rd</sup> Edition.