

Miranda Judd
7605 S. Arbory Lane
Laurel, MD 20707
mmkjudd@gmail.com
410-707-4009

EDUCATION

University of Maryland, College Park, MD *2014-2018*

Current GPA: 3.475

Major: Microbiology

Applications and Research Laboratory, Ellicott City, MD *2012-2014*

Career Academy: Biotechnology

PROFESSIONAL EXPERIENCE

University of Maryland Medical Center **Research Assistant** *June 2018 – Present*

Department of Anesthesiology

- Creation of protocols and lab procedures
- Collection and analysis of blood and urine samples
- Coordination of administrative and lab systems
- Patient interaction; management and consenting
- Management of laboratory materials and ordering
- Reagent preparation; general lab duties

Supervisors: Kenichi Tanaka, MD; LaToya Stubbs, MS

Delwiche Lab, College Park MD **Research Assistant** *March 2016 – May 2018*

Geneology of Life (GoLife) funded by NSF: Focused on the geneology of the SAR taxa

- Deconvolution microscopy of dinoflagellate species
- Staining of protists for fluorescent and light microscopy
- Single-celling protists from environmental samples
- Culturing of dinoflagellate and algal species
- Observation and analysis of culture health and characteristics via microscopy
- Microbiological media and reagent preparation; general lab duties

Supervisors: Charles Delwiche, PhD; Brittany Ott, PhD

USDA-ARS, Beltsville MD **Intern** *Sept. 2013 - Sept.2014*

*Research Project: Development and characterization of *Calonectria pseudonaviculata* mutant strains*

- Development of *Calonectria pseudonaviculata* mutants through ultra violet and N-methyl-N'-nitro-N-nitrosoguanidine random mutagenesis in order to identify phenotypic changes.
- DNA extractions and DNA sequencing; gel electrophoresis.
- Phenotypic characterization and experimentation of *C. pseudonaviculata* mutant strains testing their response to stress conditions and level of pathogenicity.
- Microbiological media and reagent preparation, general lab duties

Supervisors: Martha Malapi-Wight, PhD; Yazmin Rivera, PhD; Jo Anne-Crouch, PhD

SKILLS

Lab techniques: Chromatography, Antibody/Antigen Testing, Gel Electrophoresis, Bacterial Transformation (Chemical/ Heat Shock, Electroporation), PCR, Restriction Digest, Plant Virus Inoculations, RNA and DNA extraction, RT-PCR, DAS-ELISA, Western Blot, PAGE Electrophoresis, Restriction Mapping (Plasmids), UV-Mutagenesis, Light and Deconvolution (Delta-Vision) Microscopy, Gram-Staining of Bacteria, Culturing of Fungi, Dinoflagellates, and Algal isolates, Single-Celling Dinoflagellates, Blood and urine handling, Thrombin Generation Assay, *in vivo* Thrombus Formation assay.

TRAINING

How Flow Works, Flow Cytometry Lecture, University of Maryland, Baltimore, MD, June 3, 2019.
Introduction to Clinical and Translational Research, University of Maryland, Baltimore, MD, August 20, 2018.
HIPAA course, University of Maryland Medical Center, Baltimore, MD, June 28, 2018.
CITI Program course, University of Maryland Baltimore, Baltimore, MD, June 28, 2018.
DeltaVision Deconvolution Microscope Training. Delwiche Lab, College Park, MD, August 7, 2017

HONORS

Appleman-Norton Award for Outstanding Student in Plant Science 2018
University of Maryland, Cell Biology and Molecular Genetics Department

Office of Sustainability Mini-Grant 2016
University of Maryland, College Park

Deans List of Outstanding Students 2015-2018
University of Maryland, College Park

APS-Potomac Division, Research Poster Presentation Award 2014
Annapolis, MD

AP Scholar Award 2014
National Award

PUBLICATIONS

Judd M, Strauss E, Hasan S, Abuelkasem E, Li J, Deshpande S, Mazzeffi MA, Ogawa S, Tanaka KA. Retrospective cohort study of clinical rotational thromboelastometry results compared to in vitro cytochalasin D effects. *Journal of Cardiothoracic and Vascular Anesthesia*. 2019. (*Submitted*).

Tanaka KA, Bharadwaj S, Hasan S, **Judd M**, Abuelkasem E, Henderson RA, Chow JH, Williams W, Mazzeffi MA, Crimmins SD, Malinow AM. Elevated Fibrinogen, von Willebrand Factor, and Factor VIII Confer Resistance to Dilutional Coagulopathy and Activated Protein C in Normal Pregnant Women. *British Journal of Anaesthesia* 2019. (*Print*).

Mazzeffi M, Strauss E, Meyer M, Hasan S, **Judd M**, Abuelkasem E, Chow J, Nandwani V, McCarthy P, Tanaka K. Coagulation factor levels and underlying thrombin generation patterns in adult extracorporeal membrane oxygenation patients. *Anesthesia & Analgesia* 2019. (*Print*).

PRESENTATIONS

Judd M., Rivera Y., Malapi-Wight M., and Crouch J.A. 2014. Development and characterization of *Calonectria pseudonaviculata* mutant strains. American Phytopathological Society, Potomac Division, Annapolis, MD, USA. Poster presentation.

Judd M., Rivera Y., Malapi-Wight M., and Crouch J.A. 2014. Development and characterization of *Calonectria pseudonaviculata* mutant strains. USDA-ARS Poster Day. Beltsville, MD, USA. Poster presentation.

Judd M., Ndukwe P., Rivera Y., Malapi-Wight M., and Crouch J.A. 2014. Mutagenesis of *Calonectria pseudonaviculata* for phenotypic studies. Howard County Public School System STEM Fair, Columbia, MD, USA. Poster presentation.

Judd M., Ndukwe P., Rivera Y., Malapi-Wight M., and Crouch J.A. 2014. Mutagenesis of *Calonectria pseudonaviculata* for phenotypic studies. Applications and Research Laboratory Science Fair, Columbia, MD, USA. Poster presentation.

Hébert J, Malapi-Wight M, Rivera Y, Ismaiel A, Saeid N, Salgado-Salazar C, Gehesquière B, Dadzie H, **Judd M**, Huengens K, Crouch J.A. 2014. Studies on the genome scale: Next generation sequencing in the boxwood blight system. MD, USA. Oral Presentation.